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⑦ Applicant: **MEDICAL RESEARCH COUNCIL**  
20 Park Crescent  
London W1M 3AL (GB)

Waldmann, Hermann  
11 Gurney Way  
Cambridge (GB)

Clark, Michael Ronald  
108 York Street  
Cambridge CB1 2PY (GB)

⑦ Inventor: Clark, Michael Ronald  
108 York Street  
Cambridge, CB1 2PY (GB)

Riechmann, Lutz  
7084 Vista del Mar Avenue  
La Jolla California 92037 (US)

Waldmann, Herman  
11 Gurney Way  
Cambridge (GB)

Winter, Gregory Paul  
64 Cavendish Avenue  
Cambridge (GB)

⑦ Representative: Morion, Colin David et al  
Keith W Nash & Co. Pearl Assurance House 90-92 Regent  
Street  
Cambridge CB2 1DP (GB)

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⑤ Improvements relating to antibodies.

⑤ An antibody is produced, which will bind effectively with the antigen Campath-1, and which has at least one complementarity determining region of rat origin, as identified in Figure 2, which may be combined with a range of different foreign variable domain framework regions as desired, including framework regions of human origin.

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## Description

## Improvements in or relating to antibodies

## Field of the invention

5 This invention relates to antibodies.

## Background to the invention

Antibodies, or immunoglobulins, comprise two heavy chains linked together by disulphide bonds and two light chains, each light chain being linked to a respective heavy chain by disulphide bonds. The general structure of an antibody of class IgG (i.e. an immunoglobulin (Ig) of class gamma (G)) is shown schematically in Figure 1 of the accompanying drawings.

Each heavy chain has at one end a variable domain followed by a number of constant domains. Each light chain has a variable domain at one end and a constant domain at its other end, the light chain variable domain being aligned with the variable domain of the heavy chain and the light chain constant domain being aligned with the first constant domain of the heavy chain. The constant domains in the light and heavy chains are not involved directly in binding the antibody to antigen.

The variable domains of each pair of light and heavy chains form the antigen binding site. The domains on the light and heavy chains have the same general structure and each domain comprises four framework regions, whose sequences are relatively conserved, connected by three complementarity determining regions (CDRs) (see reference 11). The four framework regions largely adopt a beta-sheet conformation and the CDRs form loops connecting, and in some cases forming part of, the beta-sheet structure.

The CDRs are held in close proximity by the framework regions and, with the CDRs from the other domain, contribute to the formation of the antigen binding site.

## Summary of the invention

According to one aspect of the present invention there is provided an antibody having at least one CDR which is foreign with respect to the constant region of the antibody, said at least one foreign CDR being selected from CDRs substantially as identified in Figure 2, namely residues 31 to 35, 50 to 65 and 95 to 102 of the heavy chain and residues 24 to 34, 50 to 56 and 89 to 97 of the light chain, the antibody being capable of binding effectively to the antigen Campath-1.

The term "foreign" is used in relation to the CDR(s) and constant region to mean of different origin.

In Figure 2 and elsewhere in the specification amino acid residues are identified by the conventionally used one letter symbols, as follows:

35	Amino Acid	One-letter symbol
	Alanine	A
	Arginine	R
	Asparagine	N
40	Aspartic acid	D
	Asparagine or aspartic acid	B
	Cysteine	C
	Glutamine	Q
45	Glutamic acid	E
	Glutamine or glutamic acid	Z
	Glycine	G
50	Histidine	H
	Isoleucine	I
	Leucine	L
	Lysine	K
	Methionine	M
55	Phenylalanine	F
	Proline	P
	Serine	S
	Threonine	T
60	Tryptophan	W
	Tyrosine	Y
	Valine	V

In this specification, effective antibody-antigen binding is used to mean that antibody effects 50% binding to antigen at antibody concentrations of less than or equal to 70 ug/ml, preferably at concentrations of less than or equal to 7 ug/ml. Binding affinity may be tested by assay procedures such as are described in the Example herein, eg using Campath-1 antigen obtained from a glycolipid extract from human spleen. (ug = microgram)

Thus, a standard procedure for the extraction of glycolipids can be applied to the extraction of the Campath-1 antigen from human spleens. This standard extraction procedure involves the treatment of 1 volume of homogenised human spleen tissue with 3 volumes of water, 11 volumes of methanol and 5.4 volumes of chloroform. After mixing precipitated material is discarded and a further 3.5 volumes of water are added, followed by further mixing. The mixture is then allowed to separate into two phases, the lower chloroform-containing phase is discarded and the upper aqueous phase is concentrated to provide a crude extract of the Campath-1 antigen, which can if desired be purified further by affinity chromatography, for example using the YTH66.9 antibody referred to hereinafter.

The antibody of the present invention desirably has a light chain with at least one CDR selected from CDRs substantially as identified in Figure 2 and a heavy chain with at least one CDR selected from CDRs substantially as identified in Figure 2.

As a further possibility, the antibody of the present invention preferably has three heavy chain CDRs substantially as identified in Figure 2, or three light chain CDRs substantially as identified in Figure 2. More preferably, the antibody has all six heavy and light chain CDRs substantially as identified in Figure 2.

Hence, in a preferred aspect the present invention provides an antibody having heavy and light chain CDRs which are foreign with respect to the constant region of the antibody, said CDRs being substantially as identified in Figure 2, namely residues 31 to 35, 50 to 65 and 95 to 102 of the heavy chain and residues 24 to 34, 50 to 56 and 89 to 97 of the light chain, the antibody being capable of binding effectively to the antigen Campath-1.

The CDRs identified in Figure 2 are of rat origin and may be combined with a range of different variable domain framework regions, as desired, including, eg, framework regions of rat or human origin.

In a further aspect the present invention provides an antibody having heavy and light chain variable domains as identified in the lower lines of sequence information in Figure 2, namely residues 1 to 113 of the heavy chain and residues 1 to 108 of the light chain, the CDRs and constant region of the antibody being foreign with respect to one another, the antibody being capable of binding effectively to the antigen Campath-1.

Such an antibody comprises CDRs and framework regions of rat origin.

The invention also provides an antibody having heavy and light chain variable domains as identified in the upper lines of sequence information in Figure 2, namely residues 1 to 113 of the heavy chain and residues 1 to 108 of the light chain, and that will bind effectively to the antigen Campath-1.

Such an antibody comprises CDRs of rat origin in framework regions of human origin.

Such an antibody may be modified by having a phenylalanine group at residue 27 of the heavy chain in place of serine, and possibly also by having a threonine group at residue 30 of the heavy chain in place of serine. A Ser(27) to Phe mutation is found to increase antibody-antigen binding significantly. However, the mutation of Ser(30) to Thr (in the human framework with the Ser(27) to Phe mutation) has little effect on binding affinity. This illustrates that point mutations in the antibody may have a major effect or little effect on the affinity of the antibody for the antigen. Although the two changes Ser(27) to Phe and Ser(30) to Thr are located within the framework region as defined in reference 11, they lie within the hypervariable loop H1 as defined in reference 18. It is accordingly believed that some changes in the CDRs may similarly be made without necessarily having an adverse effect on antibody-antigen affinity. References to CDRs substantially as identified in Figure 2 are accordingly intended to include within their scope not only CDRs identical to those identified in Figure 2 but also variants of such CDRs, subject to the requirement of the antibody binding effectively to Campath-1.

The antibody is preferably in biologically pure form, desirably being at least 95% (by wt) free of other biological materials.

The remainder of the antibody, namely the heavy and light chain constant domains and possibly also variable domain framework regions and one or more CDRs, can be based on antibodies of various different types as desired including, eg, rat and human antibodies of different classes. Thus, the constant domains can be selected to have desired effector functions appropriate to the intended use of the antibody. For example, for therapeutic purposes, human IgG1 and rat IgG2b are currently favoured isotypes. Further, of the human IgG isotypes, IgG1 and IgG3 appear to be the most effective for complement and cell mediated lysis, and therefore for killing tumour cells. For other purposes other isotypes may be favoured, eg, rat IgM, IgG1, IgG2a, IgG2c, human IgG2, IgG4 etc. For human therapy it is particularly desirable to use human isotypes, to minimise antioglobulin responses during therapy.

The Campath-1 antigen is strongly expressed on virtually all human lymphocytes and monocytes, but is absent from other blood cells including the hemopoietic stem cells, the antigen being described by Hale et al in Blood, 1983, 62, 873-882 (reference 6). That paper describes the antibody YTH66.9 which is specific for the Campath-1 antigen, this antibody being available from Serotec of 22 Bankside, Station Approach, Kidlington, Oxford, England, under the designation YTH 66.9 HL. A further series of antibodies to Campath-1 have been produced, including rat monoclonal antibodies of IgM, IgG2a, and IgG2c isotypes (reference 7) and more recently IgG1 and IgG2b isotypes have been isolated as class switch variants from the IgG2a secreting cell line YTH 34.5HL (reference 8). All of these antibodies with the exception of the rat IgG2c isotype are able to efficiently lyse human lymphocytes with human complement.

In addition, the IgG2b antibody YTH 34.5HL-G2b, but not the other isotypes, is effective in antibody dependent cell mediated cytotoxicity (ADCC) with human effector cells (reference 8). These rat monoclonal antibodies have found important application in the context of immunosuppression, for control of graft-versus-host disease in bone marrow transplantation (reference 6); the management of organ rejection (reference 9); the prevention of marrow rejection and in the treatment of various lymphoid malignancies (reference 10). For in-vivo use, the IgG2b antibody YTH 34.5HL-G2b seems to be the most effective at depleting lymphocytes, but the use of any of the antibodies in this group is limited by the antiglobulin response which can occur within two weeks of the initiation of treatment (reference 10).

Antibodies in accordance with the invention, particularly those based on human isotypes, thus have good therapeutic potential. In particular, the availability of a reshaped human antibody with specificity for the Campath-1 antigen should permit a full analysis of the in vivo potency and immunogenicity of an anti-lymphocyte antibody with wide therapeutic potential. Such reshaped antibodies have been used in the treatment of patients with non-Hodgkin lymphoma, as well as in the treatment of some cases of autoimmune disease. Further trials with organ graft patients, particularly kidney graft patients, are proposed. Even if anti-idiotypic responses are eventually observed, considerable therapeutic benefit could be derived by an extended course of treatment. In addition an antiglobulin response restricted to idiotypic should be circumvented by using a series of antibodies with different idiotypic (reference 20). In principle, the idiotypic of the reshaped Campath-1 could be changed by altering the hypervariable regions or the framework regions: evidence from a reshaped antibody specific for the hapten nitrophenyl acetate suggests that the recognition by anti-idiotypic antisera and anti-idiotypic monoclonal antibodies is influenced by residues in the framework region (reference 5). Thus recycling the hypervariable regions on different human framework regions should change the idiotypic, although ultimately it could focus the response directly onto the binding site for Campath-1 antigen. Although such focussing would be undesirable for Campath-1 antibodies, it could be an advantage for the development of anti-idiotypic vaccines.

In further aspect, the invention thus provides a method of treating patients, particularly humans, with cancers, particularly lymphomas, or for immunosuppression purposes, comprising administering antibodies in accordance with the invention.

Antibodies in accordance with the present invention may be formulated for administration to patients by mixing antibody purified in conventional manner with a physiologically acceptable diluent or carrier, possibly in admixture with other therapeutic agents such as other antibodies. In one example, purified antibody was reconstituted in a commercially available human plasma protein solution (minus gamma globulin). The formulation was administered by intravenous infusion at the rate of 5 mg antibody per day for at least 10 days.

Antibodies in accordance with the invention can be produced in various different ways, as will be described in greater detail in the Examples following.

Heavy and light chain variable domains are conveniently produced separately and assembled with the remainder of an antibody of desired origin, eg desired human isotype.

Genes encoding the variable domains of an antibody of desired structure may be produced, and attached to genes encoding the constant domains of an antibody of desired isotype. Genes encoding variable domains can be derived from hybridoma cDNA or from the chromosome. Genes encoding the variable domains are also conveniently constructed by gene synthesis techniques or by site directed mutagenesis using long synthetic oligonucleotides. Expression is conveniently effected by transforming a cell line, eg an immortalised mammalian cell line such as a myeloma cell line, with expression vectors including DNA coding for the variable domains and for the remainder of the antibody and culturing the transformed cell line to produce the desired antibody.

In another aspect the invention provides a process for the preparation of an antibody having at least one CDR (complementarity determining region) which is foreign with respect to the constant region of the antibody, said at least one foreign CDR being selected from CDRs substantially as identified in Figure 2, that is amino acid residues 31 to 35, 50 to 65 and 95 to 102 of the heavy chain as shown in Figure 2a, and amino acid residues 24 to 34, 50 to 56 and 89 to 97 of the light chain as shown in Figure 2b, the antibody being capable of binding effectively to the antigen Campath-1, which process comprises culturing a cell capable of expressing the antibody in order to effect expression thereof.

It will be appreciated that the antibody may be used in a form which retains the CDRs but lacks other parts of the whole molecule not essential to its binding efficacy, in particular a F(ab')<sub>2</sub> fragment, and the word antibody is used herein to include such materials.

The invention will be further described, by way of illustration, in the following Examples which refer to the accompanying drawings, in which:

Figure 1 is a schematic diagram illustrating the structure of an IgG molecule;

Figure 2 illustrates nucleic acid and amino acid sequences of the variable domains of antibodies in accordance with the invention, with Figure 2a representing the heavy chain and Figure 2b representing the light chain. The upper line of the Figure gives sequence information for reshaped antibody, with the lower line giving sequence information for rat YTH 34.5HL antibody;

Figure 3 illustrates the sequence of the HuVLLYS<sup>o</sup> gene and derived amino acid sequence;

Figure 4 illustrates the sequence of the HuVLLYS gene and derived amino acid sequence;

Figure 5 illustrates a strategy for producing a reshaped human antibody having rat CDRs;

Figure 6 illustrate loop Phe 27 to Tyr 35 in the heavy chain variable domain of the human myeloma

protein KOL;

Figure 7 illustrates the results of complement lysis and ADCC for various antibodies;

Figure 8 illustrates the results of complement lysis and ADCC of various further antibodies;

Figure 9 shows the effect of CAMPATH-1H on blood counts in a patient (patient 1), with solid triangles showing results for lymphocytes and empty triangles results for neutrophils;

Figure 10 shows the cytology of bone marrow cells from two (patients 1 and 2) patients treated with CAMPATH-1H:

A = patient 1 trephine before treatment with CAMPATH-1H

B = patient 1 trephine on day 43 (ie 16 days after treatment)

C = patient 2 aspirate before treatment with CAMPATH-1H

D = patient 2 aspirate on day 78 (ie 35 days after treatment);

Figure 11 shows computed tomography scans from patients 1 and 2, showing affected spleens and lymphonode:

A = patient 1 before treatment with CAMPATH-1H

B = patient 1 on day 57

C = patient 2 before treatment with CAMPATH-H (retrocrural node arrowed)

D = patient 2 on day 51; and

Figure 12 shows the effect of CAMPATH-H on blood counts in patient 2, with solid triangles showing results for lymphocytes and empty triangles results for neutrophils.

#### Example 1

The sequences of the heavy and light chain variable domains of rat IgG2a Campath-1 antibody YTH 34.5HL were determined by cloning the cDNA (Figure 2), and the hypervariable regions were identified according to Kabat (see reference 11). Sequence information is given in the lower lines of Figure 2, with the CDRs identified in boxes.

In the heavy chain variable domain there is an unusual feature in the framework region. In most known heavy chain sequences Pro(41) and Leu(45) are highly conserved: Pro(41) helps turn a loop distant from the antigen binding site and Leu(45) is in the beta bulge which forms part of the conserved packing between heavy and light chain variable domains (reference 12). In YTH 34.5HL these residues are replaced by Ala(41) and Pro(45), and presumably this could have some effect on the packing of the heavy and light chain variable domains.

Working at the level of the gene and using three large mutagenic oligonucleotides for each variable domain, in a single step the hypervariable regions of YTH 34.5HL were mounted on human heavy or light chain framework regions taken from the crystallographically solved proteins NEW for the heavy chain (reference 13) and from a protein based closely on the human myeloma protein REI for the light chain (reference 14). The NEW light chain was not used because there is a deletion at the beginning of the third framework region of the NEW light chain. The resulting reshaped heavy chain variable domain HuVHCAMP is based on the HuVHNP gene (references 1, 5) with the framework regions of human NEW alternating with the hypervariable regions of rat YTH 34.5HL. There are discrepancies involving the first framework region and the first hypervariable loop of the NEW heavy chain between the published sequence used here and the sequence deposited in the Brookhaven data base (in parentheses): Ser27 (to Thr), Thr28 (to Ser) and Ser30 (to Asp). Neither version is definitive and the discrepancies do not affect present considerations. The reshaped light chain variable domain HuVLCAMP is a similar construct, except with essentially the framework regions of the human myeloma protein REI, with the C-terminal and the 3' non-coding sequence taken from a human J<sub>κ</sub>-region sequence (reference 22). Sequence information for the variable domain of the reshaped antibody is given in the upper lines in Figure 2. The sequences of oligonucleotide primers are given and their locations on the genes are also marked in Figure 2.

Considering the above in further detail, mRNA was purified (reference 23) from the hybridoma clone YTH 34.5HL (gamma 2a, k<sup>o</sup>), and first strand cDNA made by priming with oligonucleotides complementary to the 5' end of the CH1 (oligonucleotide I) and the Cκ exons (oligonucleotide II). cDNA was cloned and sequenced as described in references 24 and 25.

For expression of the rat heavy chain variable domain RaVHCAMP, two restriction sites (XbaI and SalI) were introduced at each end of the cDNA clone in M13 using mutagenic oligonucleotides III and V respectively, and the XbaI-SalI fragment excised. Simultaneously, the corresponding sites were introduced into the M13-HuVHNP gene using oligonucleotides IV and VI, and the region between the sites exchanged. The sequence at the junctions was corrected with oligonucleotides VII and VIII, and an internal BamHI site removed using the oligonucleotide IX, to create the M13-RaVHCAMP gene. The encoded sequence of the mature domain is thus identical to that of YTH 34.5HL.

The reshaped heavy chain variable domain (HuVHCAMP) was constructed in an M13 vector by priming with three long oligonucleotides simultaneously on the single strand containing the M13-HuVHNP gene (references 1, 5). The mutagenesis techniques used were similar to those described in reference 33, using the host 71-18 mutL and without imposing strand selection. Each oligonucleotide (X, XI and XII) was designed to replace each of the hypervariable regions with the corresponding region from the heavy chain of the YTH 34.5HL antibody.

Colony blots were probed initially with the oligonucleotide X and hybridisation positives were sequenced: the overall yield of the triple mutant was 54%. Ser27 to Phe and Ser27 to Phe, Ser30 to Thr mutants (to be described below) of M13mp8-HuVHCAMP were made with the mixed oligonucleotide XIII.

The reshaped light chain variable domain (HuVLCAMP) was constructed in an M13 vector from a gene with framework regions based on human REI. As above, three long oligonucleotides (XIV, XV, and XVI) were used to introduce the hypervariable regions of the YTH 34.5HL light chain.

Construction of the humanised light chain variable domain is described in greater detail in the following seven paragraphs.

(1) The "humanised" light chain variable domain (HuVLCAMP) was constructed in three stages, utilising a "humanised" light chain variable domain (HuVLLYS) which had been constructed for other purposes.

(a) The first stage involved the gene synthesis of a "humanised" light chain variable domain gene (HuVLLYS<sup>o</sup>). The HuVLLYS<sup>o</sup> gene incorporates human framework regions identical to the most common residue in each position in the Kabat alignment of the human kappa subgroup I, except for residues 97-108, which were identical to those in the human J1 fragment described in reference 34. The sequences of the framework regions are very similar to the crystallographically solved light chain structure REI. The CDRs in HuVLLYS<sup>o</sup> were identical to those in the mouse antilysozyme antibody (D1.3) light chain (unpublished). A 30 bp sequence, identical to the sequence following the genomic J1 segment, was introduced to the 3' side of residue 108. BamHI and EcoRI restriction sites were introduced at the 3' end of the synthetic gene, and a PstI site at the 5' end. The gene synthesis of HuVLLYS<sup>o</sup> is described in paragraphs (2) to (5) below, and the sequence of the gene and the derived amino acid sequence is given in Figure 3.

(b) The second stage involved the introduction of the HuVLLYS<sup>o</sup> gene in place of the heavy chain variable domain in the vector M13-MOVHNP and this is described in paragraphs 6 and 7 below. Thus the light chain variable domain utilises the promoter and signal sequence of a heavy chain variable domain: at the 3' end of the gene the sequence is derived from the human light chain J1 segment as described in paragraph (1a). The sequence of the HuVLLYS gene and the derived amino acid sequence is given in Figure 4.

(c) The third stage involved the conversion of HuVLLYS to a "humanised" light chain variable domain with the CDRs of Campath-1 specificity.

2. For the synthesis of the HuVLLYS<sup>o</sup> gene, three blocks of oligonucleotides (PK1-5, KK1-5 and KE1-8 in the table in paragraph 3 below) were cloned separately into M13 vectors, and sequenced. Each cloned block was excised and ligated together into M13mp19 to create the HuVLLYS<sup>o</sup> gene.

3. Oligonucleotides listed below were produced on an Applied Biosystems 380B synthesizer. Each oligonucleotide was size-purified, 10 nmol being subjected to electrophoresis on a 20 x 40 cm 12% polyacrylamide, 7M urea gel, eluted from the gel by dialysis against water, and lyophilized. For gene synthesis or mutagenesis, a 50 pmol aliquot of each purified oligonucleotide was phosphorylated in a 20 ul reaction mixture with 50mM Tris-Cl (pH 8.0), 10mM MgCl<sub>2</sub>, 5mM dithiothreitol, 1 mM ATP, and 5 units of polynucleotide kinase, incubated at 37° for 30 minutes. When used as hybridization probes, gel-purified oligonucleotides were phosphorylated in a similar fashion, except on a 15 pmol scale with an excess of <sup>32</sup>P labeled ATP.

sequence (5'-3')

GACATCCAGATGACCCAGAGCCCAAGCAGCCTGAGCGCCAGCGTGGGT	5
GACAGAGTGACCATCACCTGTAGAGCCAGCGGTAACATCCACAACCTAC	
CTGGCTTGGTAC	
CAAGCCAGGTAGTTGTGGATGTTACCGCTGGCTCTACAGGTGAT	10
GGTCACTCTGTCACCCACGCTGGCGCTCAGGCT	
GCTTGGGCTCTGGGTCATCTGGATGTCTGCA	
	15
CAGCAGAAGCCAGGTAAGGCTCCAAAGCTGCTGATCTACTACACCACC	
A	
CCCTGGCTGACGGTGTGCCAAGCAGATTCAGCGGTAGCGGTAGCGGTA	20
C	
CGCTACCGCTACCGCTGAATCTGCT	
TGGCACACCGTCAGCCAGGGTGGTGGTGTAGTAGATCAGC	25
AGCTTTGGAGCCTTACCTGGCTTCTGCTGGTAC	
	30
CGACTTCACCTTCACCATCAGCAGCCTCCAGCCAGAGGACATCGCCAC	
CTACTACTGCC	
AGCACTTCTGGAGCACCCCAAGGACGTTTCGGCCAAGGGACCAAGGTGG	35
A	
AATCAAACGTGAGTAGAATTTAACTTTGCTTCCTCAGTTGGATCCTA	
G	
KE4 AATTCTAGGATCCAACTGAGGAAGCAAAGTTTAAA	40
KE5 TTCTACTCACGTTTGATTTCACCTTGGTCCCTT	
KE6 GGCCGAACGTCCCTTGGGGTGCTCCAGAAGTGCTGGCAGTAGTAG	
KE7 GTGGCGATGTCCCTCTGGCTGGAGGCT	45
KE8 GCTGATGGTGAAGGTGAAGTCGGTAC	
	50
PK0 TCATCTGGATGTCTGGAGTGGACACCT	
	55
	60

4. The construction of individual blocks is described for the PK1-5 block, but KE1-5 and KE1-8 blocks were cloned as KpnI-KpnI and KpnI-EcoRI fragments respectively in a similar way. 4ul portions of each oligonucleotide PK1, PK2, PK3, PK4 and PK5, kinased as in paragraph 3, were combined and annealed at 80°C for 5 minutes, 67°C for 30 minutes, and allowed to cool to room temperature over the span of 30 minutes. 0.1ul

65

of this annealing mix was ligated with 20 ng of PstI/KpnI digested M13-mp19, in 10 $\mu$ l 50mM Tris-Cl (pH7.5), 10mM MgCl<sub>2</sub>, 10mM dithiothreitol, 1 mM ATP, 120 units T4 DNA ligase (Biolabs), and incubated 12 hours at 15°C. The ligation mix was used to transfect competent *E. coli* strain BMH 71-18, plated with BCIG and IPTG, and a clone containing the complete PstI-KpnI insert was identified.

5 The three cloned blocks were excised from 10 $\mu$ g double-stranded replicative form of the three M13 vectors, by digestion with PstI/KpnI (block PK1-5), KpnI (block KKI-5) and KpnI/EcoRI (block KE1-8). The inserts were separated from the vector by electrophoresis on a 20 x 20 cm 12% polyacrylamide gel, eluted from the gel slices with 0.5 M NH<sub>4</sub>OAc, 10 mM Mg (OAc)<sub>2</sub>, 0.1 mM EDTA, 0.1% SDS, and purified by phenol extraction and ethanol precipitation. All three fragments were ligated to PstI/EcoRI cut M13-mp19. 200 white  
10 plaques were transferred by toothpick to a fresh 2xTY plate, and grown as a grid of infected colonies. The plate was blotted with nitrocellulose filters, which were then treated with 0.5 M NaOH, followed by 1M Tris-Cl (pH7.5), and baked 1 hr at 80°C under vacuum. The filters were washed at 67°C in 3x Denhardt's solution, 2xSSC, 0.07% SDS, followed by 6xSSC at room temperature. Filters were then probed with the radiolabeled oligonucleotides KK3 or KK4 in 3ml of 6xSSC at 37°. Following hybridization with both oligonucleotides, positive  
15 colonies were picked for DNA sequencing. A phage clone containing correctly assembled blocks was designated M13-HuVLLYS\*.

5. To introduce the HuVLLYS\* gene in place of the heavy chain variable domain in the vector M13-MOVHNP (described in reference 5) as MV<sub>HP</sub> with HindIII site at the 3' end of the reading frame), double-stranded replicative form DNA of phages M13-HuVLLYS\* and M13-MOVHNP were prepared and digested with PstI and BamHI. The insert of M13-HuVLLYS\* was isolated on a polyacrylamide gel, and the vector portion of  
20 M13-MOVHNP was isolated on an agarose gel. The purified fragments were ligated and transfected into *E. coli* strain BMH71-18, and the resulting plaques probed with oligonucleotide KK3 to identify the insert. The clone was designated M13-HuVLLYS\*.

7. In the M13-HuVLLYS\* gene, to join the signal sequence of MOVHNP correctly to the 5' end of the  
25 HuVLLYS\* gene (at the PstI site), single stranded DNA was prepared and altered by oligonucleotide directed mutagenesis with the primer PKO- see paragraph (3) for sequence. The mutant clone was designated M13-HuVLLYS.

The reshaped human heavy and light chain variable domains were then assembled with constant domains in three stages as illustrated in Figure 5. In Figure 5 sequences of rat origin are marked in black, and those of human origin in white. The recombinant heavy and light chains are also marked using a systematic  
30 nomenclature.

The illustrated procedure permits a step-wise check on the reshaping of the heavy chain variable domain (stage 1), the selection of the human isotype (stage 2), and the reshaping of the light chain variable domain and assembly of human antibody (stage 3). The vector constructions were genomic, with the variable domains  
35 excised from the M13 vectors and cloned as HindIII-BamHI fragments and the constant domains as BamHI-BamHI fragments in either pSVgpt (heavy chain) (reference 15) or pSVneo (light chain) (reference 16) vectors. The heavy chain enhancer was included to the 5' side of the variable domain, and expression of both light and heavy chains was driven from heavy chain promoter and the heavy chain signal sequence.

The human gamma 1 (reference 26), gamma 2 (reference 27), gamma 3 (reference 28), gamma 4 (reference  
40 21) and K (reference 22) constant domains, and the rat gamma 2b (reference 29) constant domains were introduced as BamHI-BamHI fragments. The following plasmids were constructed and transfected into lymphoid cell lines by electroporation (reference 30). In stage 1, the pSVgpt vectors HuVHCAMP-RaIgG2B, and also two mutants for reasons to be explained below, HuVHCAMP(Ser27 to Phe)-RaIgG2B, HuVHCAMP(Ser27 to Phe, Ser30 to Thr)-RaIgG2B were introduced into the heavy chain loss variant of  
45 YTH34.5HL. In stage 2, the pSVgpt vectors RaVHCAMP-RaIgG2B, RaVHCAMP-HuIgG1, RaVHCAMP-HuIgG2, RaVHCAMP-HuIgG3, RaVHCAMP-HuIgG4 were transfected as described above. In stage 3, the pSVgpt vector HuI(Ser27-Phe, Ser30-Thr)VHCAMP-HuIgG1 was cotransfected with the pSV-neo vector HuVLCAMP-HuIgK into the rat myeloma cell line YO (Y B2/3.0 Ag 20) (ref. 17). In each of the three stages, clones resistant to mycophenolic acid were selected and screened for antibody production by ELISA assays. Clones secreting  
50 antibody were subcloned by limiting dilution (for YO) or the soft agar method (for the loss variant) and assayed again before 1 litre growth in roller bottles.

#### Heavy chain variable domain

In stage 1, the reshaped heavy chain variable domain (HuVHCAMP) was attached to constant domains of  
55 the rat isotype IgG2b and transfected into a heavy chain loss variant of the YTH34.5 hybridoma. The loss variant carries two light chains, one derived from the Y3 fusion partner (reference 17). The cloned rat heavy chain variable domain (RaVHCAMP) was also expressed as above.

Antibodies were harvested at stationary phase and concentrated by precipitation with ammonium sulphate, followed by ion exchange chromatography on a Pharmacia MonoQ column. The yields of antibody were  
60 measured by an ELISA assay directed against the rat IgG2b isotype, and each adjusted to the same concentration (reference 21).

The HuVHCAMP and RaVHCAMP antibodies - all of the rat IgG2b isotype - were compared in a direct binding assay to the Campath-1 antigen (obtained from a glycolipid extract from human spleen), and also in complement lysis of human lymphocytes. For measuring the binding to antigen, the partially purified  
65 Campath-1 antigen was coated onto microtitre wells. Bound antibody was detected via a biotin labelled anti-rat



IgG2b monoclonal antibody (reference 21), developed with a streptavidin-peroxidase conjugate (Amersham plc). Complement lysis of human lymphocytes with human serum as the complement source was as described in reference 7. For both binding and complement assays, the titres for the antibodies were determined by fitting the data to a sigmoid curve by a least squares iterative procedure (reference 7), and the concentration of antibody giving 50% maximal binding or lysis was noted.

The results are given in Table 1.

Table 1

Reshaping the heavy chain variable domain

heavy chain variable domain	Concentration of antibody in ug/ml at 50% binding or lysis	
	antigen binding	complement lysis
RaVHCAMP	0.7	2.1
HuVHCAMP	27.3	(*)
HuVHCAMP (Ser27 to Phe)	1.8	16.3
HuVHCAMP (Ser27 to Phe, Ser30 to Thr)	2.0	17.6

Compared with the original rat antibody, or the engineered equivalent, the antibody with the reshaped heavy chain domain HuVHCAMP bound poorly to the Campath-1 antigen and was weakly lytic. This suggested an error in the design of the reshaped domain.

There are several assumptions underlying the transfer of hypervariable loops from one antibody to another, and in particular that the antigen binds mainly to the hypervariable regions. These are defined as regions of sequence (reference 11) or structural (reference 18) hypervariability, and the locations of hypervariable regions are similar by either criterion, except for the first hypervariable loop of the heavy chain. By sequence the first hypervariable loop extends from residues 31 to 35 (reference 11) and by structure from residues 26 to 32 (reference 18). Residues 29 and 30 to form part of the surface loop, and residue 27 which is phenylalanine or tyrosine in most sequences including YTH34.5HL, helps pack against residues 32 and 34.

By way of illustration, see Figure 6 which illustrates loop Phe27 to Tyr35 in the heavy chain variable domain of the human myeloma protein KOL which is crystallographically solved (reference 31). The backbone of the hypervariable region according to Kabat (reference 11) is highlighted, and a 200% van der Waal surface is shown around Phe27 to show the interactions with Tyr32 and Met34 of the Kabat hypervariable region. In the rat YTH34.5HL heavy chain, these three side chains are conserved, but in HuVHCAMP, Phe27 is replaced by Ser: this is because, unlike most human heavy chains, in NEW the phenylalanine is replaced by serine, which would be unable to pack in the same way as phenylalanine. To restore the packing of the loop, a Ser(27) to Phe mutation was made in HuVHCAMP, and also a double mutation Ser(27) to Phe, Ser(30) to Thr (as mentioned above).

The two mutants showed a significant increase in binding to CAMPATH-1 antigen and lysed human lymphocytes with human complement. See the results given in Table 1. Thus the affinity of the reshaped antibody could be restored by altering the packing between the hypervariable regions and the framework by a single Ser(27) to Phe mutation. This suggests that alterations in the "Kabat" framework region can enhance the affinity of the affinity of the antibody, and extends previous work in which an engineered change in the hypervariable region yielded an antibody with increased affinity (reference 19).

Heavy chain constant domains

In stage 2 (Figure 5), the rat heavy chain variable domain was attached to constant domains of the human isotypes IgG1, 2, 3, and 4, and transfected into the heavy chain loss variant of the YTH34.5 hybridoma.

Antibody was harvested from cells in stationary phase, concentrated by precipitation with ammonium sulphate and desalted into phosphate buffered saline (PBS). Antibodies bound to the Campath-1 antigen coated on microtitre plates, were assayed in ELISA directed against the rat k light chain (reference 21), and adjusted to the same concentration. The antibodies were assayed in complement lysis (as described above) and ADCC with activated human peripheral blood mononuclear cells (references 21, 32). Briefly,  $5 \times 10^4$  human peripheral blood cells were labelled with  $^{51}\text{Cr}$  and incubated for 30 minutes at room temperature with different concentrations of antibody. Excess antibody was removed and a 20 fold excess of activated cells added as

effectors. After 4 hours at 37°C death was estimated by  $^{51}\text{Cr}$  release.

The results are shown in Figure 7, in which the results for rat heavy chain variable domain attached to different human isotypes are represented as follows:

- 5 IgG1 empty squares  
IgG2 empty circles  
IgG3 solid squares  
IgG4 empty triangles

10 Results of lysis with the antibody YTH34.5HL are represented by solid circles.

In complement lysis (Figure 7a), the human IgG1 isotype proved similar to the YTH34.5HL-G2b, with the human IgG3 isotype less effective. The human IgG2 isotype was only weakly lytic and the IgG4 isotype non-lytic. In ADCC (Figure 7b) the human IgG1 was more lytic than the YTH34.5HL-G2b antibody. The decrease in lysis at higher concentration of the rat IgG2b and the human IgG1 antibody is due to an excess of antibody, which causes the lysis of effector cells. The human IgG3 antibody was weakly lytic, and the IgG2 and IgG4 isotypes were non-lytic.

The human IgG1 isotype was therefore suitable for a reshaped antibody for therapeutic use. Other recent work also suggests the IgG1 isotype as favoured for therapeutic application. When the effector functions of human isotypes were compared using a set of chimaeric antibodies with an anti-hapten variable domain, the IgG1 isotype appeared superior to the IgG3 in both complement and cell mediated lysis (reference 4). Furthermore, of two mouse chimaeric antibodies directed against cell surface antigens as tumour cell markers, with human IgG1 or IgG3 isotypes, only the IgG1 isotype mediated complement lysis (references 2, 3).

#### 25 Light chain

In stage 3 (Figure 5), the reshaped heavy chain was completed, by attaching the reshaped HuVHCAMP (Ser27 to Phe, Ser30 to Thr) domain to the human IgG1 isotype. The reshaped light chain domain HuVHCAMP was attached to the human Ck domain. The two vectors were cotransfected into the non-secreting rat Y0 myeloma line.

30 Antibody HuVHCAMP (Ser27 to Phe, Thr30 to Ser)-HuIGG1, HuVLCAMP-HuIGK was purified from supernatants of cells in stationary phase by affinity chromatography on protein A Sepharose. The antibody was at least 95% (by wt) pure. The yield (about 10mg/l) was measured spectrophotometrically. Complement and ADCC assays were performed as described in connection with Figure 7.

35 The results are shown in Figure 8, in which the results are reshaped human antibodies are represented by squares and those for rat YTH34.5HL antibodies are represented by solid circles.

The purified antibody proved almost identical to the YTH34.5HL-G2b antibody in complement lysis (Figure 8a). In cell mediated lysis the reshaped human antibody was more reactive than the rat antibody (Figure 8b). Similar results to the ones in Figure 8b were obtained with three different donors of target and effector cells (data not shown). Furthermore the antibody was as effective as YTH34.5HL-G2b in killing leukaemic cells from three patients with B Cell lymphocytic leukaemia by complement mediated lysis with human serum.

40 The rat antibody and fully humanised antibody were compared in a direct binding assay to Campath-1 antigen. Antibody concentrations were determined as described in Figures 7 and 8. The amount of rat antibody bound to partially purified Campath-1 antigen was determined as described in connection with Table 1. The amount of human antibody bound was determined by an ELISA assay using a biotinylated sheep anti-human IgG antibody (Amersham).

Table 2

#### 50 Reshaping the heavy and light chain variable domains simultaneously

antibody	Concentration of antibody in ug/ml at 50% binding antigen binding
RaVHCAMP Ra1GG2B	
RaVHCAMP RaKappa	1.01
HuVHCAMP (Ser27 to Phe, Ser30 to Thr)	
Hu1GG1	
HuVLCAMP HuKappa	1.11

65 Thus by transplanting the hypervariable regions from a rodent to a human antibody of the IgG1 subtype, the

antibody can be reshaped for therapeutic application.

The strategy illustrated in Figure 5 is stepwise assembly to allow any problems to be detected at each stage (reshaping of heavy chain variable domain, selection of constant domain and reshaping of light chain variable domain). It is quite possible to build the reshaped antibody in a single step assembly, i.e. constructing the two reshaped variable domains, attaching to appropriate constant domains and cotransfecting into e.g. YO.

## Example 2

### Patients and Methods

Antibody HuVHCAMP (Ser 27 to Phe, Thr 30 to Ser) - HuIGG1, HuVLCAMP - HuIGK, hereinafter referred to as CAMPATH-1H, was prepared as described in Example 1. The CAMPATH-1H antibodies were obtained from culture supernatant of cells growing in a hollow fibre bioreactor ('Acusyst - Jr', Endotronics) and purified by affinity chromatography on protein-A-'sepharose'. They were dissolved in phosphate-buffered saline, sterile filtered, and tested for pyrogen and sterility. Patients were prehydrated over night and antibody, diluted in 500 ml saline, was infused over 2-4 hours.

Campath-1 expression on tumour cells was measured by flow cytometry and complement-mediated lysis (references 6, 35). Serum concentrations of CAMPATH-1H were measured by immunofluorescence with normal lymphocytes. Southern blot analysis with an immunoglobulin J $\kappa$  probe was used to detect residual tumour cells in DNA extracted from mononuclear fractions of bone marrow. Antiglobulin responses were sought by two techniques. The first was a solid-phase enzyme-linked assay using microtitre plates coated with CAMPATH-1H. After incubation with patients' serum samples, the assay was developed with biotin-labelled CAMPATH-1H followed by streptavidin-peroxidase. A mixture of monoclonal mouse antibodies against human IgG was used as a positive control and 500 ng/ml of this mixture would be detected. In the second assay, patients serum samples were mixed with red cells coupled with CAMPATH-1H (reference 10). Agglutination by 5 ng/ml of the control mixture would be detected. Immunoglobulin allotypes were determined by means of standard reagents and techniques from the Central Laboratory of the Netherlands Red Cross Blood Transfusion Service.

## RESULTS

### Patient 1

A 69-year-old woman presented in 1983, and grade 1, stage IVA non-Hodgkin lymphoma in leukaemic phase was diagnosed. Between 1983 and 1988 the patient received various types of treatment, including chemotherapy and radiotherapy and rat antibody to Campath-1. She was then given treatment with CAMPATH-1H.

The starting dose was 1 mg daily and, since it was well tolerated, the dose was increased to a maximum of 20 mg/day, though the usual was 4 mg/day owing to the small amount available. In all the patient received 126 mg over 30 days. The response was prompt; in 6 days night sweats had abated, by day 10 there was pronounced reduction in splenomegaly and recovery of blood neutrophils, and by day 18 lymphoma cells were cleared from the blood (Figure 9).

On day 28 a bone marrow aspirate and trephine were hypocellular but showed active myelopoiesis and erythropoiesis and no lymphoid cells (Figure 10B). No CAMPATH-1 positive cells could be detected by flow cytometry. DNA from the mononuclear marrow cells was germline when probed with an immunoglobulin J $\kappa$  probe under conditions where clonal rearrangements could be detected in 0.2% of cells. Thus, it is concluded that lymphoma cells were cleared from the marrow. The spleen volume was reduced about eight-fold (Figures 11A, B), although it was still slightly larger than normal.

Other than fever occurring about 1 hour after the end of antibody infusions there were no adverse effects of antibody treatment until the 5th week, when severe rigors occurred after each infusion. No antiglobulin response could be detected and the rate of clearance of antibody from the serum was unchanged. For the next 3 weeks the patient continued to experience occasional fever and rigors. She was given oral cotrimoxazole because of her lymphopenia, but no infective cause of these symptoms could be found.

In the next 4 months lymphocytes, which appeared morphologically normal, slowly reappeared in the blood (up to  $0.2 \times 10^9/l$ ). They did not show the characteristic rearranged immunoglobulin fragments, and both CD3-positive and CD19-positive cells were present (see Table 3). Serum immunoglobulin levels, which had been very low since presentation, have risen towards normal (Table 3). A marrow aspirate and trephine taken 50 days after the end of treatment were again hypocellular but had no lymphomatous infiltration. This marrow sample contained 4% CAMPATH-1-positive cells and showed some oligoclonal rearrangements of immunoglobulin genes. However, by day 100, lymphoma cells were again detected in the blood and the spleen size had started to increase. A second course of 12 days' therapy with CAMPATH-1H was completed with similar therapeutic benefit to the first and no adverse effects. Since the main reservoir of disease in the patient appeared to be the spleen, splenectomy was carried out at the end of this second course of treatment. At that time no tumour cells could be detected in blood or marrow. The patient is now well 37 days after the splenectomy. The lymphocyte count is low but she has normal neutrophil, platelet, and red-cell counts.

Patient 2

A 67-year old man presented in April 1988 with splenic pain; there was 12 cm splenomegaly, and computed tomography scan of thorax and abdomen revealed retrocrural and para-aortic lymphadenopathy, the largest node measuring 3 cm in diameter (Figure 11C). A blood count revealed  $36.6 \times 10^9$  lymphocytes/ml, the majority being lymphoplasmacytoid cells which expressed surface IgG-kappa and were characterised by large cytoplasmic periodic-acid-Schiff-positive vacuoles which could be intensely stained by anti-IgG. A marrow aspirate contained 72% lymphomatous cells (Figure 10C). DNA from blood mononuclear cells showed biallelic rearrangement of immunoglobulin J<sub>H</sub> genes but was germline with various T-cell receptor and oncogene probes. The lymphoma cells expressed the CAMPATH-1 antigen in amounts comparable with normal lymphocytes but were more resistant to complement-mediated lysis. Stage IVA grade I lymphoplasmacytoid non-Hodgkin lymphoma was diagnosed.

The patient was offered CAMPATH-1H as primary therapy and received a total of 85 mg over 43 days. This resulted in clearance of the lymphoma cells and normal lymphocytes from blood (Figure 12) and marrow (Figure 10D), resolution of splenomegaly, and improvement in the lymphadenopathy. A computed tomography scan taken 8 days after the end of antibody treatment was normal (Figure 11D). A bone marrow aspirate taken at the same time showed active haemopoiesis but no lymphoma cells, and no CAMPATH-1-positive cells could be detected by flow cytometry. DNA from the mononuclear fraction of this marrow showed only germline configuration when probed with the immunoglobulin J<sub>H</sub> probe. By day 78 morphologically normal blood lymphocytes began to reappear and none of the vacuolated cells could be seen. The patient remains well and off therapy.

Some toxic effects of CAMPATH-1H were observed. The first dose was stopped after 3 mg had been given because of hypotension, possibly caused by tumour lysis. This problem was subsequently avoided by giving smaller doses at a slower rate and when lymphoma cells had been cleared from the blood, the dose was increased to a maximum of 8 mg over 4 h without any effect on blood pressure. Nevertheless, all doses induced fever (up to 38.5°C), and malaise for up to 36 h, but these were not severe enough to stop antibody treatment which, after the first week, was given on an outpatient basis. Treatment was stopped after 43 days because of the development of an urticarial rash after two successive antibody infusions.

Half-life of CAMPATH 1-H

The concentration of CAMPATH-1H was measured in serum samples taken before and after antibody infusions and at other times throughout treatment. In theory, a dose of 4-6 mg corresponds to about 1 µg/ml in the plasma. In fact free antibody could not be detected until day 4-6, about 5-20% after 24 hours.

Lack of Antiglobulin Response

The allotype of CAMPATH-1H is G1m(1,2,17),Km(3). Patient 1 was G1m(1,3,17),Km(3) and patient 2 was G1m(3),Km(3), so both could theoretically have made an anti-allotype response as well as a response to the hypervariable regions. However, we failed to detect any antiglobulin to CAMPATH-1H either by the solid-phase enzyme-linked assay or by the more sensitive haemagglutination assay. In addition, the rate of clearance of CAMPATH-1H did not change and free antibody could be detected for up to 8 days after the last dose had been given. It is possible that the reactions experienced at the end of the course of treatment could have been provoked by contaminating non-human proteins in the antibody preparation.

Discussion

The remissions achieved in these two patients show that it is possible to clear large numbers of tumour cells with small amounts of an unmodified monoclonal antibody. The effects in the first patient were far superior to the results of the previous chemotherapy and radiotherapy. The selective lysis of lymphoma cells with recovery of normal haemopoiesis during the course of treatment was an important advantage, which allowed treatment to be given largely on an outpatient basis. It is believed the choice of antibody-specificity and isotype is important; indeed, it may be why these tests had more success than previous efforts with other monoclonal antibodies. (References 36-38.) The CAMPATH-1 antigen seems to be a good target because it is widely distributed and abundant, and does not suffer from antigenic modulation. (References 6,35.)

TABLE 3 - PATIENT CHARACTERISTICS BEFORE AND AFTER TREATMENT WITH CAMPATH-1H

		Case 1		Case 2	
Diagnosis		Stage IVB grade I NHL in leukaemic phase		Stage IVA grade I lympho- plasmacytoid NHL	
		before	after	before	after
Spleen size	ml	4460	590	2600	440
Lymphadenopathy		nil	nil	retrocrural paraortic	nil
Bone marrow					
lymphoma cell %		99	0	72	0
Southern blot analysis					
Ig J <sub>H</sub> fragment		R/R	G/G	R/R	G/G
Peripheral blood					
haemoglobin	g/dl	8.7	10.6	11.2	12.0
reticulocytes	x10 <sup>9</sup> /l	31	135	nd	nd
platelets	x10 <sup>9</sup> /l	37	50	110	453
lymphocytes	x10 <sup>9</sup> /l	60	0	37	0
neutrophils	x10 <sup>9</sup> /l	0	2.0	4.6	7.3

	monocytes	x10 <sup>9</sup> /l	0	0.04	1.5	0.5
5	<hr/>					
	Lymphocyte phenotype					
	CD19	%	97	46	93	<5
10	CD3	%	0	32	8	80
	CAMPATH-1M	%	96	nd	95	nd
15	CAMPATH-1H	%	98	nd	97	nd
	<hr/>					
	Serum immunoglobulins					
20	IgM	g/l	<0.3	1.2	<0.3	0.7
	IgA	g/l	<0.5	<0.5	<0.5	0.5
25	IgG	g/l	5.8	8.2	3.2	4.7
	Bence-Jones		nil	nil	++	nil
30	<hr/>					

35 The post-therapy values refer to measurements made shortly after the end  
of antibody therapy, except for lymphocyte phenotyping and serum Ig, which  
were assessed 6 weeks later. Lymphocyte phenotypes were measured by  
40 immunofluorescence and APAAP techniques. Spleen and lymph node dimensions  
were measured by serial CT scanning (Fig11).  
45 nd = not done

50

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#### Claims

1. An antibody having at least one CDR (complementarity determining region) which is foreign with respect to the constant region of the antibody, said at least one foreign CDR being selected from CDRs substantially as identified in Figure 2, that is amino acid residues 31 to 35, 50 to 65 and 95 to 102 of the heavy chain as shown in Figure 2a, and amino acid residues 24 to 34, 50 to 56 and 89 to 97 of the light chain as shown in Figure 2b, the antibody being capable of binding effectively to the antigen Campath-1. 45
2. An antibody as claimed in claim 1, having a heavy chain with at least one CDR selected from CDRs substantially as identified in Figure 2a and a light chain with at least one CDR selected from CDRs substantially as identified in Figure 2b. 50
3. An antibody as claimed in claim 1, having a heavy chain with three CDRs substantially as identified in Figure 2a or a light chain with three CDRs substantially as identified in Figure 2b.
4. An antibody having heavy and light chain CDRs which are foreign with respect to the constant region of the antibody, said CDRs being substantially as identified in Figure 2, that is residues 31 to 35, 50 to 65 and 95 to 102 of the heavy chain as shown in Figure 2a and residues 24 to 34, 50 to 56 and 89 to 97 of the light chain as shown in Figure 2b, the antibody being capable of binding effectively to the antigen Campath-1. 55
5. An antibody as claimed in any one of claims 1 to 4, wherein the CDRs are combined with variable domain framework regions of human origin.
6. An antibody having heavy and light chain variable domains as identified in the lower lines of sequence information in Figure 2, that is residues 1 to 113 of the heavy chain and residues 1 to 108 of the light chain, the CDRs and constant region of the antibody being foreign with respect to one another, the antibody being capable of binding effectively to the antigen Campath-1. 60
7. An antibody having heavy and light chain variable domains as identified in the upper lines of sequence information in Figure 2, namely residues 1 to 113 of the heavy chain and residues 1 to 108 of the 65

light chain, and that will bind effectively to the antigen Campath-1.

8. An antibody as claimed in claim 7, wherein the phenylalanine at residue 27 in the heavy chain is replaced by serine.

9. An antibody as claimed in claim 7 or 8, wherein the threonine at residue 30 in the heavy chain is replaced by serine.

10. An antibody as claimed in any one of the preceding claims, wherein the heavy and light chain constant domains are of human IgG1 class.

11. An antibody composition, for administration to patients, comprising an antibody as claimed in any one of the preceding claims in substantially biologically pure form, together with a physiologically acceptable diluent or carrier.

#### Claims for the following Contracting States ES,GR

1. A process for the preparation of an antibody having at least one CDR (complementarity determining region) which is foreign with respect to the constant region of the antibody, said at least one foreign CDR being selected from CDRs substantially as identified in Figure 2, that is amino acid residues 31 to 35, 50 to 65 and 95 to 102 of the heavy chain as shown in Figure 2a, and amino acid residues 24 to 34, 50 to 56 and 89 to 97 of the light chain as shown in Figure 2b, the antibody being capable of binding effectively to the antigen Campath-1, which process comprises culturing a cell capable of expressing the antibody in order to effect expression thereof.

2. A process as claimed in claim 1, for preparing an antibody having a heavy chain with at least one CDR selected from CDRs substantially as identified in Figure 2a and a light chain with at least one CDR selected from CDRs substantially as identified in Figure 2b.

3. A process as claimed in claim 1, for preparing an antibody having a heavy chain with three CDRs substantially as identified in Figure 2a or a light chain with three CDRs substantially as identified in Figure 2b.

4. A process for the preparation of an antibody having heavy and light chain CDRs which are foreign with respect to the constant region of the antibody, said CDRs being substantially as identified in Figure 2, that is residues 31 to 35, 50 to 65 and 95 to 102 of the heavy chain as shown in Figure 2a and residues 24 to 34, 50 to 56 and 89 to 97 of the light chain as shown in Figure 2b, the antibody being capable of binding effectively to the antigen Campath-1, which process comprises culturing a cell capable of expressing the antibody in order to effect expression thereof.

5. A process as claimed in any one of claims 1 to 4, for preparing an antibody wherein the CDRs are combined with variable domain framework regions of human origin.

6. A process for the preparation of an antibody having heavy and light chain variable domains as identified in the lower lines of sequence information in Figure 2, that is residues 1 to 113 of the heavy chain and residues 1 to 108 of the light chain, the CDRs and constant region of the antibody being foreign with respect to one another, the antibody being capable of binding effectively to the antigen Campath-1, which process comprises culturing a cell capable of expressing the antibody in order to effect expression thereof.

7. A process for the preparation of an antibody having heavy and light chain variable domains as identified in the upper lines of sequence information in Figure 2, namely residues 1 to 113 of the heavy chain and residues 1 to 108 of the light chain, and that will bind effectively to the antigen Campath-1, which process comprises culturing a cell capable of expressing the antibody in order to effect expression thereof.

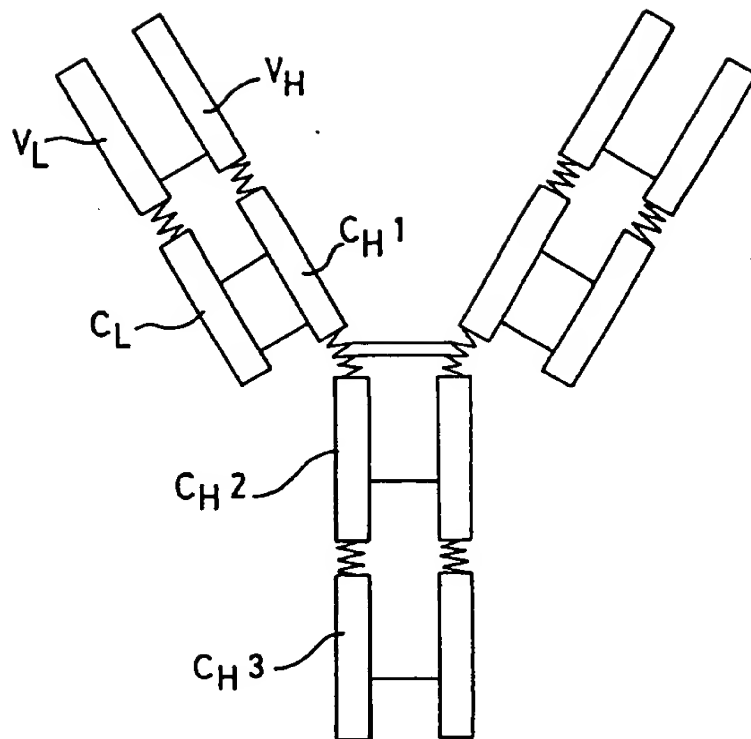
8. A process as claimed in claim 7, for preparing an antibody wherein the phenylalanine at residue 27 in the heavy chain is replaced by serine.




9. A process as claimed in claim 7 or 8, for preparing an antibody wherein the threonine at residue 30 in the heavy chain is replaced by serine.

10. A process as claimed in any one of the preceding claims, for preparing an antibody wherein the heavy and light chain constant domains are of human IgG1 class.

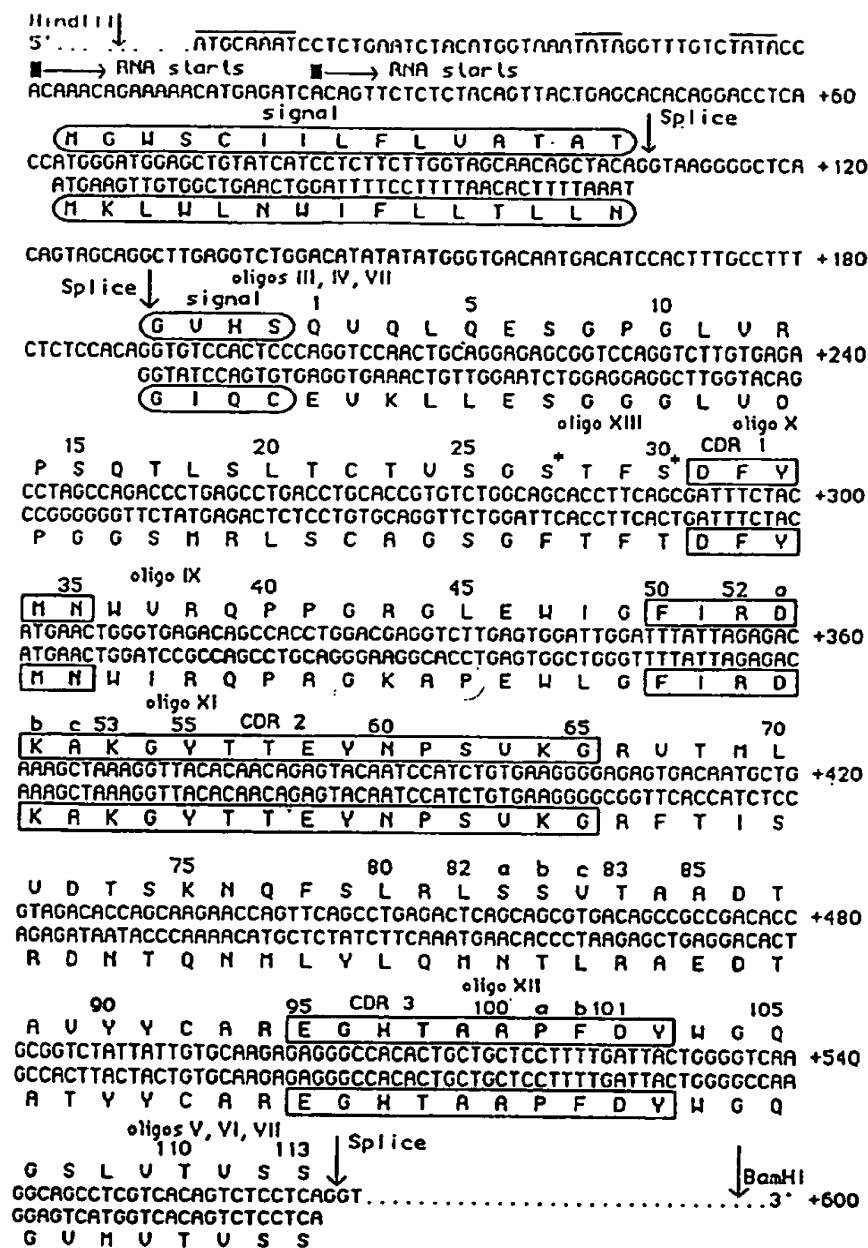
11. A process for the preparation of an antibody composition, for administration to patients, comprising mixing an antibody prepared by the process as claimed in any one of the preceding claims in substantially biologically pure form with a physiologically acceptable diluent or carrier.





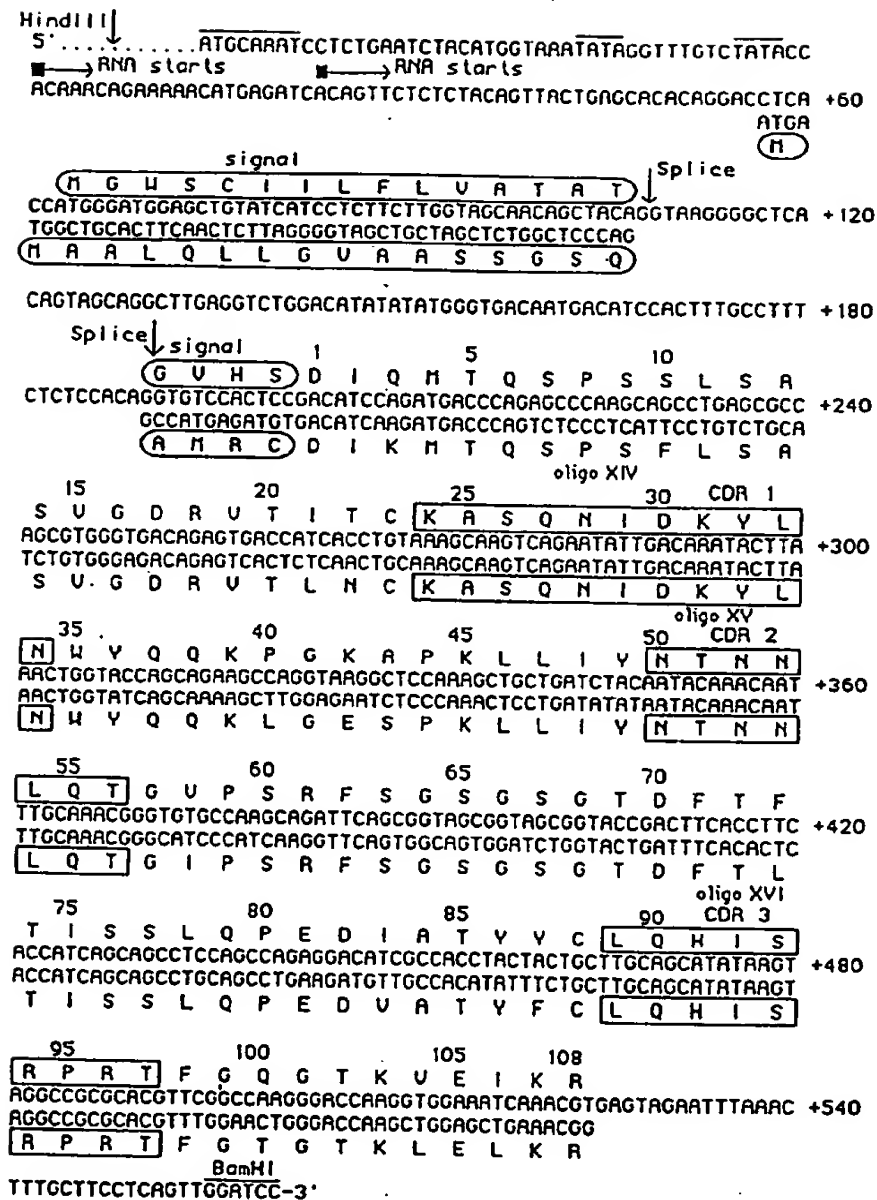
- |   |                         |
|---|-------------------------|
|  | = DOMAINS               |
|  | = INTER-DOMAIN SECTIONS |
|  | = DISULPHIDE BOND       |
| V   | = VARIABLE              |
| C   | = CONSTANT              |
| L   | = LIGHT CHAIN           |
| H   | = HEAVY CHAIN           |

*Fig.1*



Oligonucleotides: I: 5'-GGC CAG TGG ATA GAC-3', III: 5'-CAG TTT CAT CTA  
 GAA CTG GAT A-3', IV: 5'-GCA GTT GGG TCT AGA AGT GGA CAC C-3',  
 V: 5'-TCA GCT GAG TCG ACT GTG AC-3', VI: 5'-TCA CCT GAG TCG ACT GTG  
 AC-3', VII: 5'-AGT TTC ACC TCG GAG TGG ACA CCT-3', VIII: 5'-TCA CCT GAG  
 GAG ACT GTG AC-3', IX: 5'-GGC TGG CGA ATC CAG TT-3', X: 5'-CTG TCT CAC  
 CCA GTT CAT GTA GAA ATC GCT GAA GGT GCT-3', XI: 5'-CAT TGT CAC TCT  
 CCC CTT CAC AGA TGG ATT GTA CTC TGT TGT GTA ACC TTT AGC TTT GTC  
 TCT AAT AAA TCC AAT CCA CTC-3', XII: 5'-GCC TTG ACC CCA GTA ATC AAA  
 AGG AGC AGC AGT GTG GCC CTC TCT TGC ACA ATA-3', XIII: 5'-AGA AAT  
 CGG/C TGA AGG TGA AGC CAG ACA C-3'.

Fig.  
2a



Oligonucleotides: II: 5'-TGC AGC ATC AGC C-3', XIV: 5'-CTG CTG GTA CCA  
 GTT TAA GTA TTT GTC AAT ATT CTG ACT TGC TTT ACA GGT GAT GGT-3',  
 XV: 5'-GCT TGG CAC ACC CGT TTG CAA ATT GTT TGT ATT GTA GAT CAG  
 CAG-3', XVI: 5'-CCC TTG GCC GAA CGT GCG CGG CCT ACT TAT ATG CTG CAA  
 GCA GTA GTA GGT-3'.

Fig. 2b

## Sequence of the synthetic gene HUVLLYSO

D I Q M T Q S P S S L S A S V G D R V  
 CTGCA GACATCCAGATGACCCAGAGCCCAAGCAGCCTGAGCGCCAGCGTGGGTGACAGAG  
 GACGT CTGTAGGTCTACTGGGTCTCGGGTTCGTGCGACTCGCGGTGCGACCCACTGTCTC  
 10 20 30 40 50 60

T I T C R A S G N I H N Y L A W Y Q Q K  
 TGACCATCACCTGTAGAGCCAGCGGTAACATCCACAACCTACCTGGCTTGGTACCAGCAGA  
 ACTGGTAGTGGACATCTCGGTCGCCATTGTAGGTGTTGATGGACCGAACCATGGTCGTCT  
 70 80 90 100 110 120

P G K A P K L L I Y Y T T T L A D G V P  
 AGCCAGGTAAGGCTCCAAAGCTGCTGATCTACTACACCACCACCCTGGCTGACGGTGTGC  
 TCGGTCCATTCCGAGGTTTCGACGACTAGATGATGTGGTGGTGGGACCGACTGCCACAGC  
 130 140 150 160 170 180

S R F S G S G S G T D F T F T I S S L Q  
 CAAGCAGATTGAGCGGTAGCGGTACCGACTTCACCTTCACCATCAGCAGCCTCC  
 GTTCGTCTAAGTCGCCATCGCCATCGCCATGGCTGAAGTGGAGTGGTAGTCGTCGGAGG  
 190 200 210 220 230 240

P E D I A T Y Y C Q H F W S T P R T F G  
 AGCCAGAGGACATCGCCACCTACTACTGCCAGCACTTCTGGAGCACCCCAAGGACGTTG  
 TCGGTCTCCTGTAGCGGTGGATGATGACGGTCGTGAAGACCTCGTGGGGTTCCCTGCAAGC  
 250 260 270 280 290 300

Q G T K V E I K R  
 GCCAAGGGACCAAGGTGGAAATCAACGTGAGTAGAATTTAACTTTGCTTCCTCAGTTG  
 CGGTTCCCTGGTTCACCTTTAGTTTGCACATCATCTTAAATTTGAAACGAAGGAGTCAAC  
 310 320 330 340 350 360

GATCCTAGAATTC  
 CTAGGATCTTAAG  
 370

Fig.3

ATGCAAATCCTCTGAAT  
 CTACATGGTAAATATAGGTTTGTCTATAACACAAACAGAAAAACATGAGATCACAGTTCT  
 CTCTACAGTTACTGAGCACACAGGAOCTCAOCATGGGATGGAGCTGTATCATOCTCTTCT  
 M G W S C I I L F  
 L V A T A T  
 TGGTAGCAACAGCTACAGGTAAGGGGCTCACAGTAGCAGGCTTGAGGTCTGGACATATAT  
 G V H S D I Q  
 ATGGGTGACAATGACATOCACCTTTGCTTTTCTCTOCACAGGTGTCCACTCCGACATCCAG  
 5 10 15 20  
 M T Q S P S S L S A S V G D R V T I T C  
 ATGACCCAGAGCCCAAGCAGCCTGAGGCGCAGGGTGGGTGACAGAGTGACCATCACTGT  
 \*\*\*\*\*  
 25 30 35 40  
 R A S G N I H N Y L A W Y Q Q K P G K A  
 AGAGCCAGGGTAACATOCACAACCTAOCCTGGCTTGGTACCAGCAGAAGCCAGGTAAGGCT  
 \*\*\*\*\*  
 45 50 55 60  
 P K L L I Y Y T T T L A D G V P S R F S  
 CCAAGCTGCTGATCTACTACACCACCCCTGGCTGAOGGTGTGCCAAGCAGATTCAGC  
 65 70 75 80  
 G S G S G T D F T F T I S S L Q P E D I  
 GGTAGCGGTAGCGGTACCGACTTCAOCTTCAOCATCAGCAGCCTCCAGCCAGAGGACATC  
 \*\*\*\*\*  
 85 90 95 100  
 A T Y Y C Q H F W S T P R T F G Q G T K  
 GCAOCTACTACTGCCAGCACTTCTGGAGCACCCCAAGGACGTTGGGCCAAGGGAACCAAG  
 105 108  
 V E I K R  
 GTGCAAATCAAACGTGAGTAGAATTTAAACTTTGCTTCTCAGTTGGATCCTAGAATTC

Fig. 4

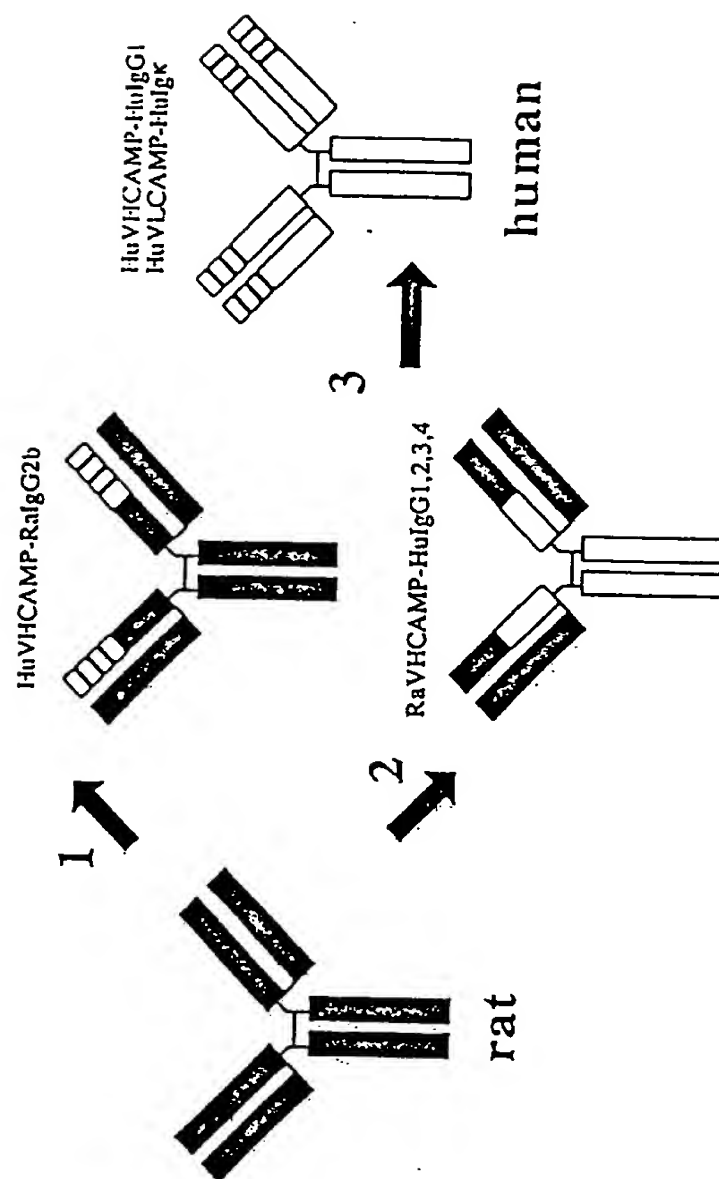


Fig. 5

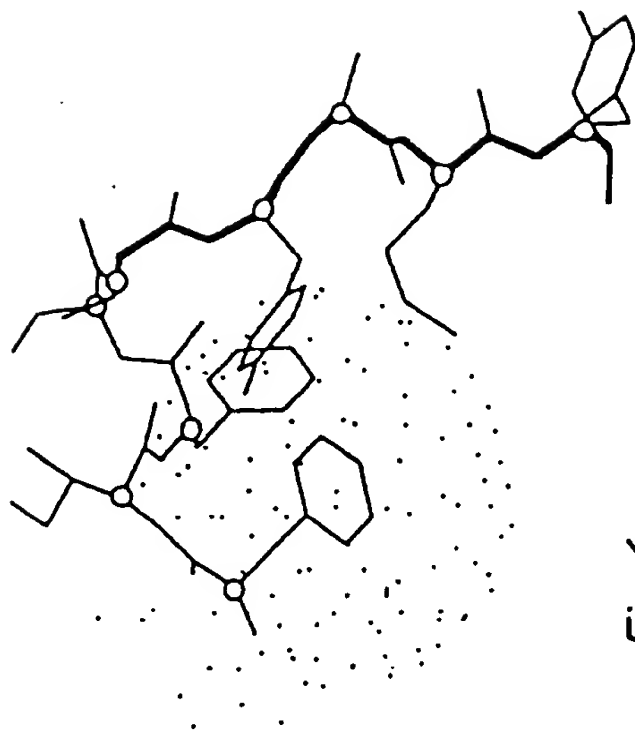
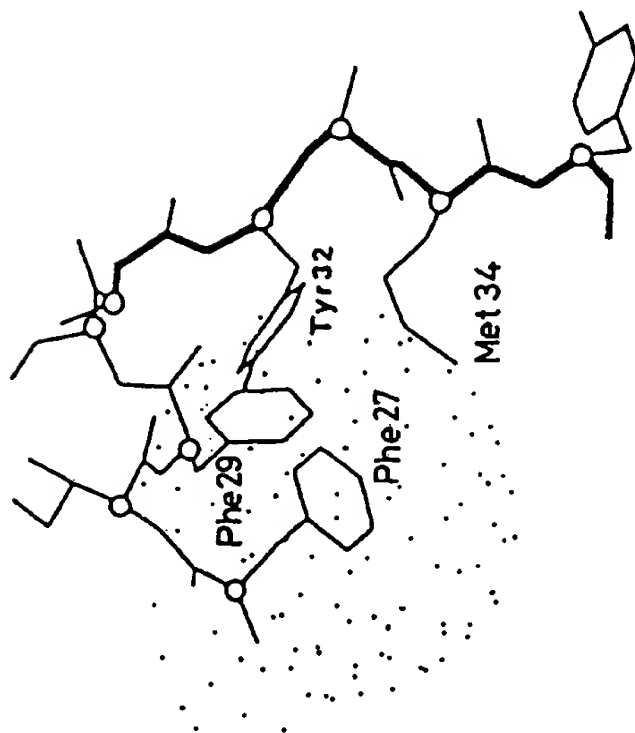
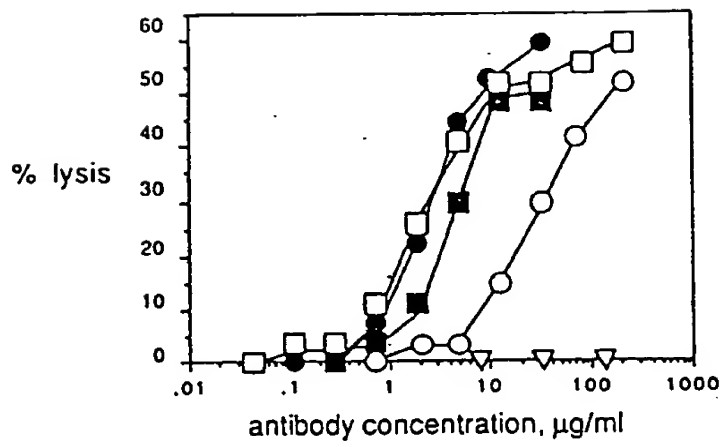
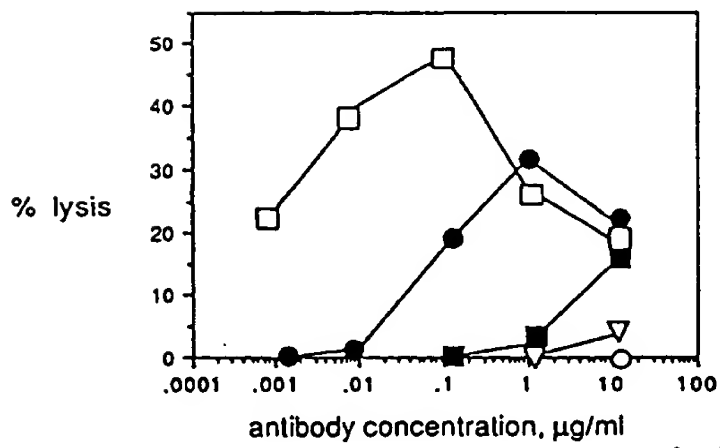


Fig.6





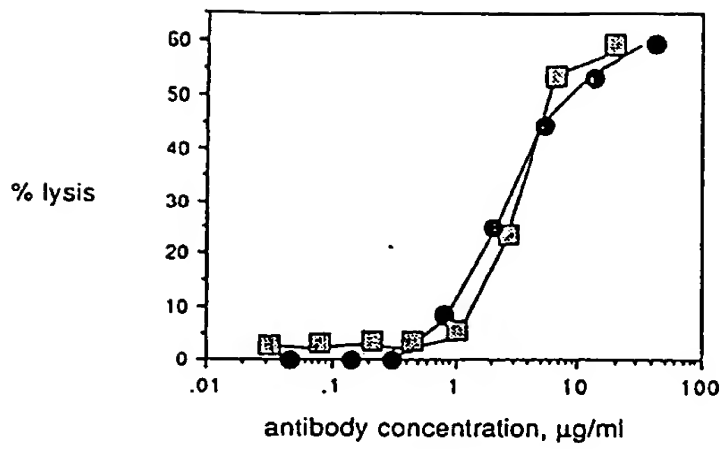
a.)



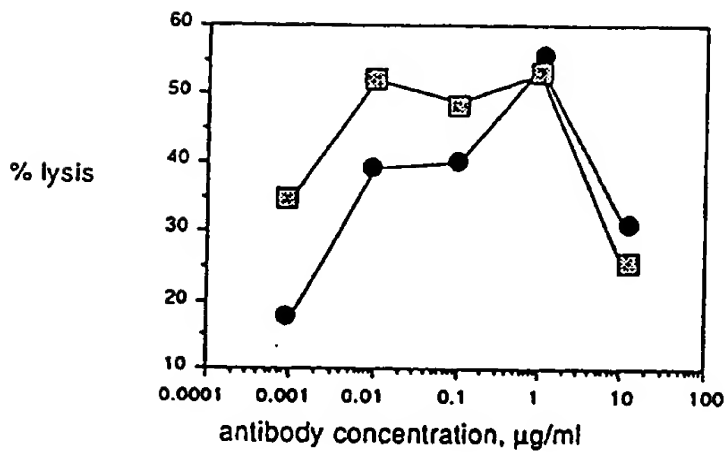
b.)

Fig. 7





a.)



b.)

Fig. 8

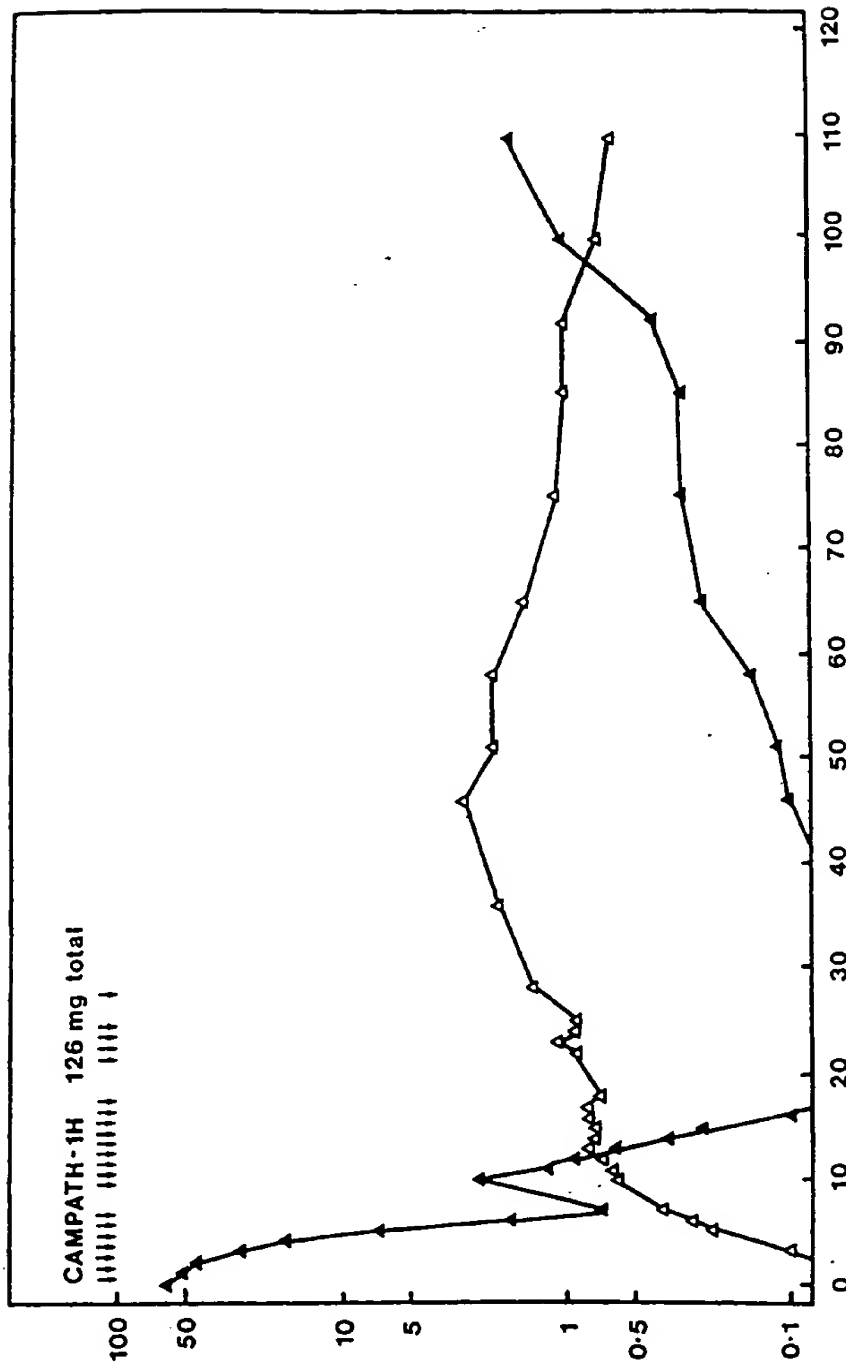


Fig.9

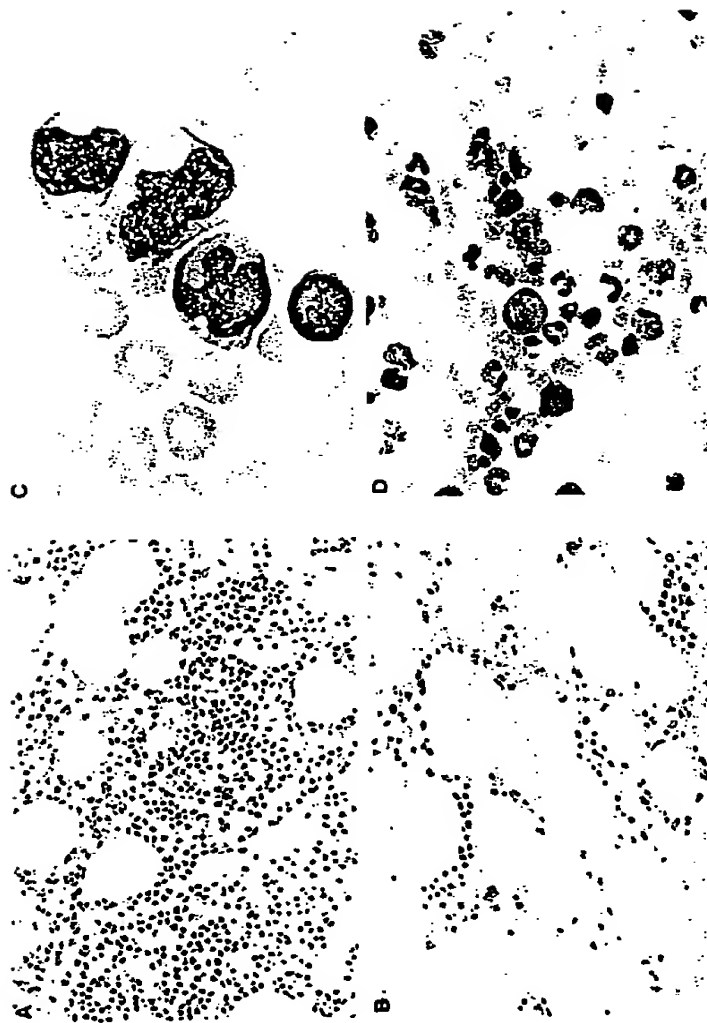


Fig. 10

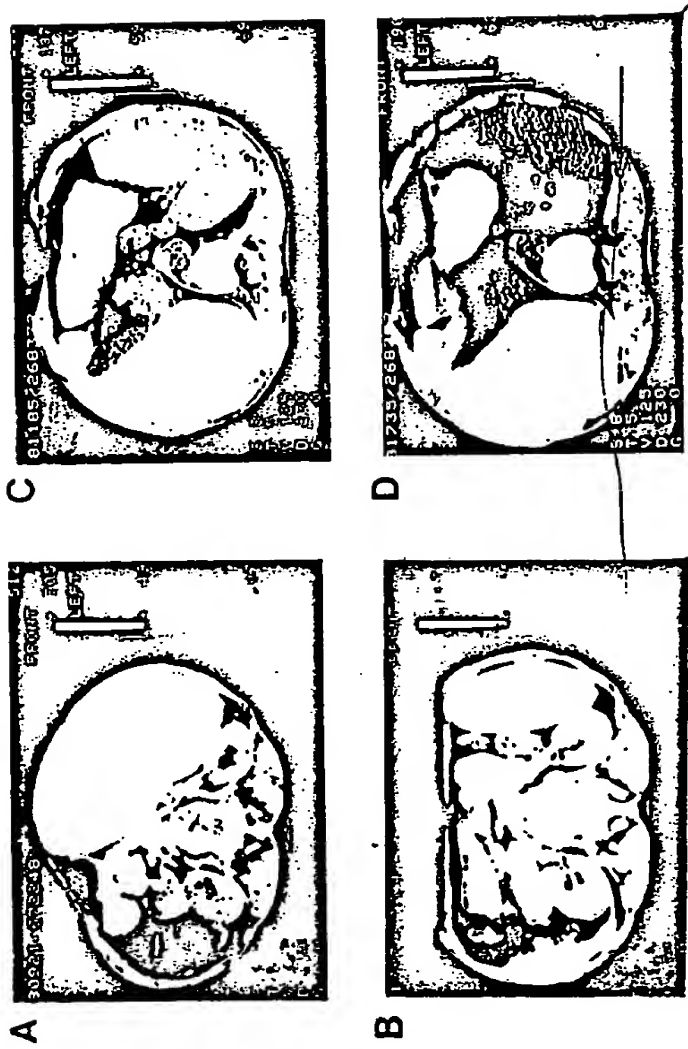


Fig.11

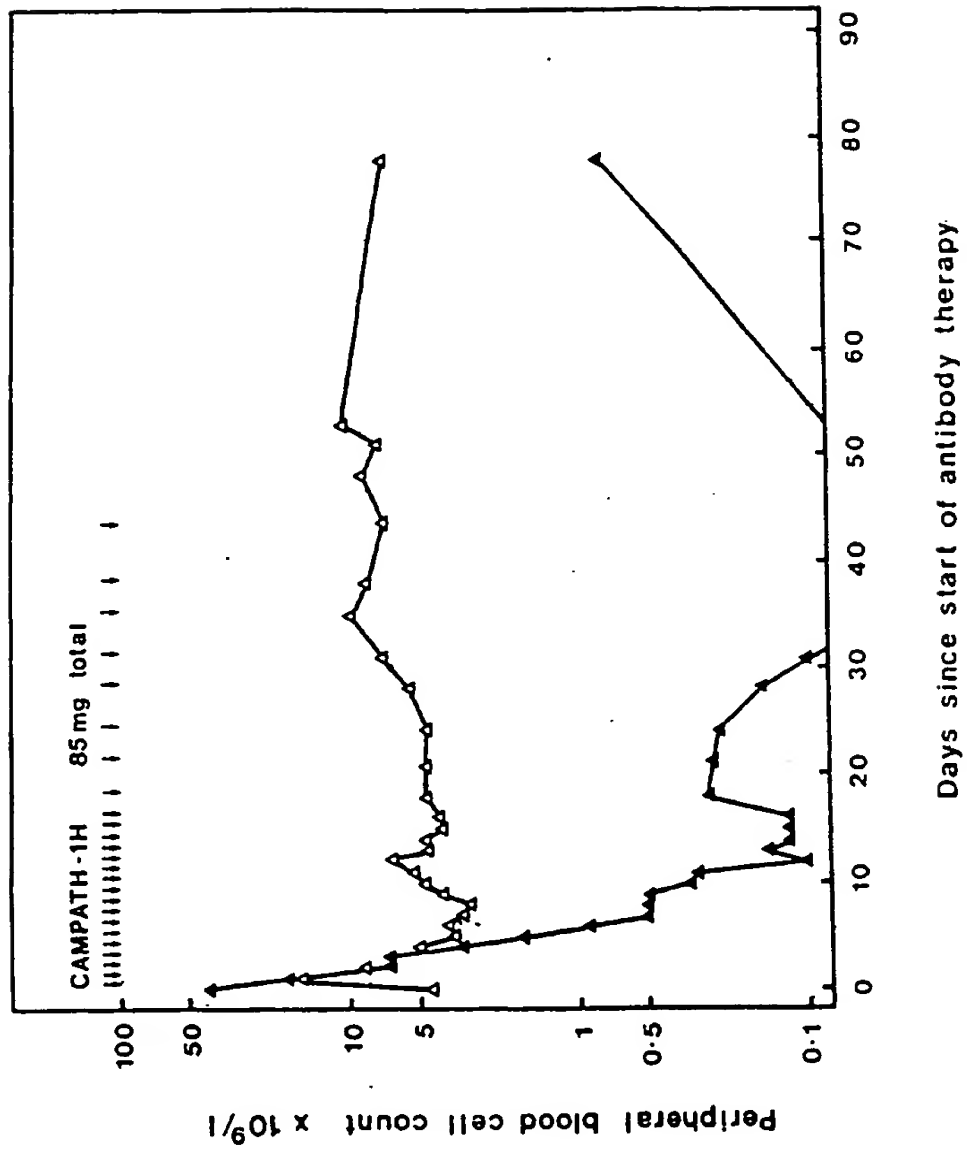


Fig.12



European Patent  
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# EUROPEAN SEARCH REPORT

Application Number

EP 89 30 1291

DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int. Cl.4)
X	EP-A-0 239 400 (G.P. WINTER) * Whole document, especially page 30, last paragraph * ---	1-11	A 61 K 39/395 C 12 N 15/00
Y,D	JOURNAL OF IMMUNOLOGICAL METHODS, vol. 103, 1987, pages 59-67, Elsevier Science Publishers B.V. (Biomedical Division); G. HALE et al.: "Isolation of low-frequency class-switch variants from rat hybrid myelomas" * Whole article * ---	1-11	
Y,D	NATURE, vol. 321, 29th May 1986, pages 522-525; P.T. JONES et al.: "Replacing the complementarity-determining regions in a human antibody with those from a mouse" * Whole article * ---	1-11	
X,P	NATURE, vol. 332, 24th March 1988, pages 323-327; L. RIECHMANN et al.: "Reshaping human antibodies for therapy" * Whole article * -----	1-11	TECHNICAL FIELDS SEARCHED (Int. Cl.4)  C 12 N C 12 P
The present search report has been drawn up for all claims			
Place of search THE HAGUE		Date of completion of the search 11-05-1989	Examiner CUPIDO M.
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